
A STUDY OF ANTIFUNGAL SUSCEPTIBILITY USING PHENOTYPIC METHOD

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ABSTRACT

Susceptibility testing of organisms against antifungal medications ordinarily utilized for treatment is a vital part of the consideration of patients with obtrusive contagious diseases. Antifungal susceptibility testing (AFST) has advanced in late a very long time to at last get normalized and accessible as both Clinical and Laboratory Standards Institute (CLSI) and European Committee on Antimicrobial Susceptibility Testing (EUCAST) reference techniques and in business manual/computerized phenotypic strategies. In clinical practice, the Sensititre Yeast One and Etest strategies are broadly utilized for AFST, especially for sterile site separates of *Candida*. All things considered, AFST is advancing toward new phenotypic strategies, for example, framework helped laser desorption ionization season of-flight mass spectrometry (MALDI-TOF MS), that are equipped for giving quick, and possibly more significant, results for the treating clinician. Our goal is to sum up refreshed information on phenotypic techniques for AFST of *Candida* and *Aspergillus* species and to survey their essentialness considering contradicting, however arising, atomic genotypic strategies.

KEYWORDS: antifungal susceptibility; *Candida*; *Aspergillus*; CLSI; EUCAST; MALDI-TOF MS

INTRODUCTION

Intrusive parasitic diseases, particularly those brought about by the species *Candida* and *Aspergillus*, keep on ascending in recurrence [1] and, alarmingly, are related with antifungal opposition [2], which makes the administration of patients with such contaminations especially testing [3,4]. Aside from contaminations because of naturally antifungal-safe species, most of these diseases are clinically treatable by three at present accessible antifungal medication classes:

triazoles (fluconazole, itraconazole, voriconazole, posaconazole, isavuconazole), echinocandins (anidulafungin, caspofungin, micafungin), and polyenes (amphotericin B-deoxycholate with its lipid and liposomal details) [5,6]. Resembling the generally late presentation of new antifungal medication classes [7] and the disclosure of novel specialists [8] for the treatment of intrusive contagious contaminations, the field of antifungal susceptibility testing (AFST) has advanced enormously in the previous quite a while [9]. Aside from handy, solid, and reproducible research facility strategies for AFST, there has been a significant push toward techniques ready to correspond in vitro lab tests with clinical result and distinguish new, clinically significant opposition systems, as has been accomplished for susceptibility testing of microscopic organisms [10]. The objective of performing AFST is to create noteworthy information for the treating clinician on the susceptibility, transitional (or portion subordinate) susceptibility, or opposition aggregate for a living being antifungal specialist blend. In a perspective article distributed ~15 years prior, Rex and Pfaller [11], while examining the precision of the "90–60 guideline" by which AFST can anticipate the result of treatment, said that AFST "has surely grown up as a helpful clinical device." This eventually worked out later through not just refinement of the Clinical and Laboratory Standards Institute (CLSI) and European Committee on Antimicrobial Susceptibility Testing (EUCAST) reference techniques as of now set up [12–15], yet additionally development of business and mechanized strategies for AFST [16]. These accomplishments improved the probability that testing a few creature drug mixes (most prominently *Candida* species and the azole antifungal specialists) could conveniently impact the choice of treatment, consequently supporting clinicians in the administration of hard to-treat parasitic diseases [17]. In the interim, better consistency and exactness of testing, alongside clinical results and pharmacokinetic/pharmacodynamic information, prompted the formation of all around approved clinical breakpoints (CBPs), in any event for azoles and regular *Candida* species [9]. Notwithstanding, in lieu of CBPs, setting up epidemiological cutoff esteems (ECVs) assisted with recognizing wild-type (WT) segregates from those that may hold a procured opposition instrument and are less inclined to react to a given antifungal specialist (non-WT) [18]. The targets of this paper are to give reports on new information from AFST examines and to talk about how AFST may improve results of obtrusive parasitic contaminations. Considering their clinical significance, we will zero in on AFST of *Candida* and *Aspergillus* species.

CONVENTIONAL PHENOTYPIC ASSAYS FOR TESTING FUNGAL SUSCEPTIBILITY

Reference AFST Methods

At present, phenotypic examines to act in vitro AFST for either yeasts or filamentous organisms (additionally named molds) incorporate two generally perceived standard techniques, CLSI [12,13] and EUCAST [14,15], which apply the stock miniature weakening strategy (BMD). Both measure antifungal action, communicated as the base inhibitory fixation (MIC) of an antifungal medication, which shows the negligible medication focus that represses contagious development. Regardless of some methodological contrasts (e.g., glucose focus, inoculum size, understanding endpoint, and so on) between the two [16], CLSI and EUCAST have been demonstrated to, endless supply of testing, similar MIC information for all classes of antifungal specialists. Specifically, the EUCAST technique utilizes a higher level of glucose (2%) in the test medium to encourage expanded parasitic development [14,15], which might be especially prompted when testing molds. To quicken the opportunity to AFST results, the two strategies incorporate (in any event for *Candida* species) a suggested brooding season of 24 h [20]. In view of various total MIC values created, EUCAST and CLSI have set up disparate CBPs (i.e., MIC limits used to arrange disengages as vulnerable or safe) for *Candida* species. As opposed to CLSI, which has not set CBPs against any molds (counting *Aspergillus* species) [23], EUCAST gives form CBPs, with species-related CBPs decided for *A. fumigatus*, *A. flavus*, *A. nidulans*, *A. niger*, and *A. terreus*. The two guidelines propose the idea of least successful fixation (MEC) for perusing echinocandin AFST consequences of molds. Notwithstanding, MEC assurance isn't in every case simple due to its dependence on surveying the progress purpose of hyphae from ordinary to distorted structures, which regularly requires tiny perception. As a rule, reference BMD tests are in fact requesting and not expected for routine lab practice. Moreover, the interlaboratory inconstancy in caspofungin MICs noted with *Candida* species may fundamentally frustrate the utilization of both CLSI and EUCAST techniques. Regardless, they hold incredible incentive as vital comparators in assessing execution investigations of business techniques, for example, those examined underneath. In one enormous examination distributed as of late, great relationship was gotten between EUCAST (EDef 7.2) and CLSI (M27-A3) for amphotericin B,

flucytosine, anidulafungin, caspofungin, micafungin, fluconazole, isavuconazole, itraconazole, posaconazole, and voriconazole among 357 disengages of *Candida* species, demonstrating >93% clear cut arrangement for all antifungal specialists tried. Low understanding principally respected testing of amphotericin B, anidulafungin, and isavuconazole against *C. glabrata*, and caspofungin against *C. parapsilosis*, *C. tropicalis*, and *C. krusei*, prompting further calls for more harmonization.

Commercial AFST Methods

As recently checked on [16], either BMD strategies, which use shading endpoints because of metabolic color (e.g., AlamarBlue) fused into development media (e.g., SensititreTMYeastOneTM (SYO; Thermo Fisher Scientific, Waltham, MA USA)), or agar-based techniques, which use focus inclinations of antifungals that diffuse into development media (e.g., Etest[®]; AB Biodisk, Solna, Sweden), are changes of the CLSI/EUCAST reference strategies. A substantial option to these (manual) examines is to perform AFST through computerized implies (e.g., VITEK[®] 2 framework; bioMérieux, Marcy-l'étoile, France). These phenotypic strategies are as such restricted by requiring an unadulterated culture of the contaminating creature prior to testing. All things considered, SYO, Etest, and VITEK 2 are business techniques generally utilized for in vitro AFST of *Candida* and additionally *Aspergillus* species, and are believed to be better than reference strategies being used, comfort, and adaptability [16]. Since its presentation in routine microbiology research facilities, the SYO microdilution antifungal board—permitting concurrent testing of amphotericin B, echinocandins, and triazoles—has been broadly assessed for yeasts, turning into the focal point of enormous emergency clinic considers.

For CLSI CBPs/ECVs to relegate susceptibility (or the WT aggregate) to foundationally dynamic antifungal specialists (the SYO's maker suggests utilizing CLSI CBPs), two ongoing examinations wrote about SYO MIC results for *Candida* species. In the investigation by Posteraro et al., susceptibility/WT rates to amphotericin B and flucytosine were over 97% in all yeast detaches (n = 1250, including *Candida* and non-*Candida* species). Rates for fluconazole (barring *C. krusei*), itraconazole, and voriconazole were 98.7% in *C. albicans*, 92.3% in the *C. parapsilosis* species intricate, 96.1% in *C. tropicalis*, 92.5% in *C. glabrata*, and 100% in both *C.*

guilliermondii and *C. krusei*. Rates for echinocandins were 99.7% to 99.8% in all *Candida* species. Essentially, Xiao et al. discovered that over 99.3% of the disengages ($n = 1072$, including all normal non-albicans *Candida* species) had a WT aggregate to amphotericin B and flucytosine. Susceptibility/WT rates for azoles among *C. parapsilosis* species complex disengages were $\geq 97.5\%$. Among $\sim 14.3\%$ of fluconazole-safe *C. glabrata* segregates, 11.6% were cross-impervious to fluconazole and voriconazole. All *C. krusei* segregates were helpless/WT to voriconazole, posaconazole, and itraconazole, though 97.7% to 100% of secludes were vulnerable to caspofungin, micafungin, and anidulafungin. Echinocandins speak to first-line treatment of intrusive *Candida* diseases. Procured echinocandin obstruction is basically seen among *C. albicans* and *C. glabrata* and is related with transformations in two limited problem area (HS) areas (HS1 and HS2) of FKS1 (*C. albicans* and *C. glabrata*) and FKS2 (*C. glabrata* just) qualities. Eschenauer et al. underscored that embracing CLSI CBPs for caspofungin may exaggerate the rates at which secludes of *C. glabrata* and *C. krusei* are nonsusceptible to caspofungin. While clinical microbiology research centers should utilize micafungin and anidulafungin as substitute markers to anticipate susceptibility or protection from caspofungin, a few creators have attempted to build up ECVs utilizing the SYO strategy. Espinell-Ingroff et al. determined SYO ECVs for echinocandins and eight *Candida* species. Outstandingly, SYO ECVs for anidulafungin, caspofungin, and micafungin effectively named non-WT 88.9% (72/81), 91.4% (74/81), and 93.8% (76/81), separately, of *Candida* secludes with known FKS changes. Regardless of their affectability for distinguishing problem area changes, the positive and negative prescient estimations of these ECVs in routine clinical application were not decided. Recently, Kritikos et al. determined echinocandin ECVs for *C. albicans* ($n = 1277$) and *C. glabrata* ($n = 347$) tried by SYO and evaluated their capacity to distinguish FKS freaks in a 10-year candidemia review from the FUNGINOS network. Among 70 detaches with $MIC \geq ECV$ for any echinocandin and afterward sequenced, no FKS transformation was found in "as far as possible WT" disconnects ($MIC = ECV$ for at any rate one echinocandin), recommending a phenomenal negative prescient estimation of these ECVs. Among the 18 "non-WT" disengages ($MIC > ECV$ for at any rate one echinocandin), FKS changes were found in the main two disconnects with $MIC > ECV$ for each of the three echinocandins, yet not in the secludes having

a "non-WT" aggregate for just a couple echinocandins. Be that as it may, approving these SYO EVCs in settings with higher paces of echinocandin opposition stays to be finished.

MALDI-TOF Mass Spectrometry-Based AFST Methods

In numerous European clinical microbiology research centers, for example, our own, the appearance of framework helped laser desorption ionization season of-flight mass spectrometry (MALDI-TOF MS) has radically modified the routine indicative work process. Accordingly, MALDI-TOF MS offers the opportunity to distinguish practically all microbial genera and species with phenomenal unwavering quality, quickness, and cost-viability. In any case, this has happened rapidly for microscopic organisms however not all that rapidly for parasites. Challenges connected to the intricacy of the parasitic cell hampered early enhancement of test expository techniques and, thus, huge scope appropriation of MALDI-TOF MS in clinical mycology. Until now, MALDI-TOF MS examination is appropriate for microbial separates refined from essential examples or positive blood societies, fundamentally lessening the turnaround time contrasted with biochemical or nucleic corrosive based procedures, for example, DNA sequencing. In any case, high MALDI-TOF MS execution, especially for secretive species inside the *Candida* and *Aspergillus* species edifices, must be accomplished with the proper information bases gave by some promoted MALDI-TOF MS frameworks. While susceptibility testing techniques are not straightforwardly pertinent to essential examples, some ongoing examinations have announced accomplishment with aggregate focused (or semimolecular) MALDI-TOF MS strategies for AFST. With regards to past work that presented the negligible profile change focus (MPCC) as another endpoint for AFST, our exploration bunch created MALDI-TOF MS-based examines for testing the echinocandin susceptibility of parasitic species. In one of the principal examines, De Carolis et al. gotten mass spectra from parasitic cells presented to various caspofungin fixations for 15 h, and afterward coordinated the "middle" mass spectra with each of the "outrageous" mass spectra utilizing composite connection list (CCI) examination. MPCC speaks to the CCI esteem at which a range is more like the range saw at the maximal caspofungin focus (greatest CCI) than the range saw at the invalid caspofungin fixation (invalid CCI).

The creators demonstrated that MPCC values approximated MIC (or MEC) values for 100% of *Candida* and *Aspergillus* strains tested (altogether, 44 among WT and FKS1 mutant strains). In the subsequent experiment, which was the first streamlined (here named MS-AFST), Vella et al. given separation among vulnerable and safe isolates of *C. albicans* after 3 h of introduction of parasitic cells at three antifungal medication levels: no medication (invalid fixation), transitional ("breakpoint"), and most extreme (maximal focus). By methods for this "three-point" measure, segregates were powerless or safe when the CCI values got by coordinating their breakpoint range with their greatest range were, separately, higher or lower than the CCI values acquired by coordinating their breakpoint range with their range at invalid focus. Utilizing this model, 100% (51/51) and 90.9% (10/11) of the disengages tested yielded MALDI AFST results that were as per the WT or FKS1-mutant genotype, separately. To broaden our discoveries, Vella et al. attempted to approve the 3 h MS-AFST measure with a board of 80 clinical isolates of *C. glabrata* tested against echinocandin (anidulafungin) and triazole (fluconazole) antifungal specialists. Albeit gained azole opposition in *Candida* species is multifaceted, the enlistment of medication efflux encoded by CDR qualities and guideline of their appearance by changes in the record factor CgPdr1 (encoded by the CgPDR1 quality) speak to the most clinically pertinent atomic instruments. The investigation uncovered that 85.0% (68/80) and 96.2% of disengages had arrangement results for anidulafungin and fluconazole that completely concurred with those acquired by the FKS1/FKS2 genotype or CgCDR1/CDR2 overexpression antifungal-obstruction systems. While examining the MS-AFST results as per the FKS1/FKS2 genotype, arrangement was 100% (6/6) for disengages with a changed FKS1 quality and 25.0% (4/16) for isolates with a transformed FKS2 quality. This brought about 15.0% of mistaken arrangements for anidulafungin that elaborate FKS2 HS1 changes. As per the CLSI reference strategy, MS-AFST tests yielded upwards of 11 significant blunders (i.e., a safe isolate misclassified as powerless) with anidulafungin, and just two significant mistakes with fluconazole. Curiously, inconsistencies could be settled with MS-AFST tests performed at 15 h of presentation to both antifungal medications; for this situation, MPCC values were incidental with the MICs for those isolates demonstrating discrepant outcomes. Taken together, these discoveries exhibit that MS-AFST in the 3 h design neglected to identify *C. glabrata* disengages with echinocandin-related FKS2 transformations.

How to Better Use Phenotypic Fungal Susceptibility Results in the Clinic Setting

Expected to dependably recognize patients whose disease is probably going to react to a given antifungal specialist, *in vitro* susceptibility testing has improved our capacity to foresee the result of treatment [9], yet is continually defied with expanding protection from antifungal specialists [3]. Since the far and wide utilization of triazoles in mid 1990, antifungal obstruction in both *Candida* and *Aspergillus* has become a genuine general medical issue. The ascent in echinocandin opposition, azole obstruction, and cross-protection from at least two antifungal classes (multidrug obstruction) in pathogenic growths has included types of *Candida*, for example, *C. glabrata* and, recently, *C. auris*. Also, the occurrence of azole-safe *A. fumigatus* has endangered results for high-hazard patients, in light of the fact that the rejection of azole antifungal medications from prophylaxis or first-line treatment of obtrusive aspergillosis would restrict drug decisions. Nonetheless, we are cognizant that different host, drug, and parasitic components add to helpful disappointments, and there is no total relationship between *in vitro* MIC and clinical reaction. This has blocked CLSI or EUCAST from building up CBPs for some antifungal specialists and parasitic species, despite the fact that both the CLSI and EUCAST techniques, which bring about various CBPs, are grounded in pharmacodynamic reactions in creature models and patients [75]. Concerning *Candida*, antifungal susceptibility is unsurprising if the tainting living being is distinguished to the species level, yet individual secludes may not follow this course, subsequently requiring antifungal testing [9]. Critically, the Infectious Diseases Society of America (IDSA) rules for the administration of candidiasis suggest regularly performing AFST for *C. glabrata* against azoles and echinocandins [5]. Similar rules notice that normal testing for *Candida* species other than *C. glabrata* has less worth [5]. By the by, we concur with the assessment by Ostrosky-Zeichner and Andes [9] that regularly testing antifungal susceptibility of all circulation system and sterile site separates of *Candida* species might be useful to give a file to susceptibility patterns and the rise of obstruction locally and provincially. Notwithstanding, in an asset confined climate, AFST should zero in on secludes from instances of treatment disappointment, advancement disease, or restricted helpful alternatives, which are outcomes of hidden comorbidities, unfriendly occasions, or past antifungal use [9]. Steady with this, in a situation of earlier echinocandin presentation, Vella et al. recommended that quick

recognition of *C. glabrata* detaches as fluconazole-safe by the MS-AFST measure could make clinicians aware of the possible presence of anidulafungin obstruction in these segregates (Figure 1). Regardless of speaking to an individual perspective on the utility of this methodology (and not a proposal), Figure 1 underlines that utilizing anidulafungin as a proxy marker would prompt a situation in which, if no obstruction is identified, both of the three echinocandins (not just anidulafungin) could be regulated.

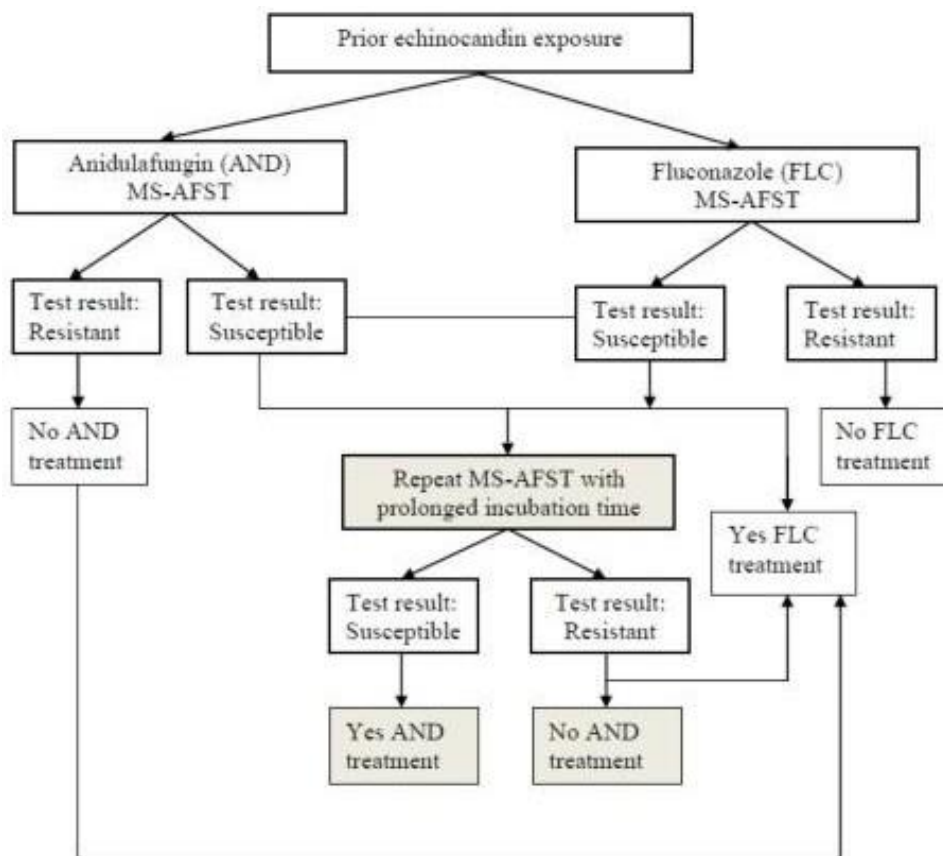


Figure 1. Potential treatment of invasive *C. glabrata* infection based on mass spectrometry–antifungal susceptibility testing (MS-AFST) results. In a clinical context of prior echinocandin exposure, results of susceptibility or resistance to anidulafungin and/or fluconazole within 3 h or, in cases of isolates with FKS2 HS1 mutations, 6–12 h after testing may guide the appropriate administration of antifungal therapy

Concerning *Aspergillus*, the IDSA rules for the administration of aspergillosis suggest performing susceptibility testing against azoles fundamentally for patients who neglect to react to treatment or for epidemiological purposes [6]. In any case, these rules don't indicate research facility testing strategies for the detachment of *Aspergillus* from respiratory plot tests. As of late, a global master board gathered to think the administration of azole-safe intrusive aspergillosis, presuming that in culture-positive cases, in vitro susceptibility testing is exceptionally shown when antifungal treatment is proposed. Up to five states should be tried in patients who are to get antifungal treatment in geographic locales with azole opposition, which is likewise suggested by the European rules for aspergillosis distributed in mid 2018 [81]. Clinically utilized triazole antifungals are subordinates of either fluconazole (voriconazole and isavuconazole) or ketoconazole (itraconazole and posaconazole) as the lead compound. This corresponds with the cross-opposition aggregates saw in clinical (and natural) *A. fumigatus* detaches with the CYP51A transformations that are inside the itraconazole/voriconazole and voriconazole/isavuconazole compound sets. In this manner, while in any event voriconazole and itraconazole are suggested as screening drugs, the VIP check (Nijmegen, The Netherlands), a business agar-based technique that comprises of four wells containing voriconazole, itraconazole, posaconazole, or a development control, was created for effectively separating between azole-helpless and - safe disconnects of *A. fumigatus*. Clinical screening considers are urged to create epidemiological information, which thus may assist with reevaluating clinical treatment alternatives on a neighborhood or public premise. In this specific circumstance, Dudakova et al. proposed a work process for assessing *A. fumigatus* secludes from such screening considers that distinguishes genuine positives and yields powerful information on the predominance and phylogenetic relatedness of safe disengages (Figure 2). In the writers' own insight, the types of each separate can be promptly decided by means of MALDI-TOF MS utilizing the business information bases. The secludes with a dull blue-green appearance, which is regular to *A. fumigatus* and its kin species, however not a dependable MALDI-TOF MS distinguishing proof as *A. fumigatus*, end up being secretive species not yet remembered for the plan introduced. Furthermore, the creators arranged the MIC values associating with singular amino corrosive replacements in the CYP51A-encoded compound for translation of DNA sequencing information, particularly without refined *A. fumigatus* detaches.

CONCLUSION

With respect to *Candida* and *Aspergillus*, the contamination related death rates are still inadmissibly high, in spite of ongoing advances in prophylaxis, early analysis, and treatment of contagious illnesses. Antifungal prophylaxis with antimold-dynamic azole mixes (posaconazole or voriconazole) to diminish the rate of obtrusive form contaminations in high-hazard patients might be related with advancement diseases brought about by uncommon multidrug-safe molds. Thusly, precise assurance of antifungal susceptibility of organisms, which may likewise incorporate non-*Aspergillus* molds (*Mucorales*, *Fusarium* spp., or *Scedosporium apiospermum* complex), is required at any rate in explicit circumstances during the consideration of patients with obtrusive parasitic contaminations. Regardless of whether the ordinary antimicrobial susceptibility tests are as yet valuable will rely upon how quick the walk toward fast phenotypic antimicrobial susceptibility testing. More work is relied upon to broaden MALDI-TOF MS–based AFST to clinically applicable parasitic microbes other than *Candida* or *Aspergillus*, in light of the fact that there are no theoretical obstacles to do this. Later on, the achievement or disappointment of recently arose innovations, as proportions of improved patient results, will rely basically upon how extraordinary the nearby predominance of antifungal obstruction is and how objectively the innovation is incorporated into the clinical microbiology lab practice.

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